

## ORIGINAL PAPER

# Influence of some agrochemical products on Loquat inflorescence blight, leaf spot and fruit rot diseases.

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## ABSTRACT

Inflorescence blight, leaf spot and fruit rot are the most important diseases that significantly decrease the production of loquat trees in Egypt. The typical symptoms were observed on loquat trees cv. Emanuel (15-years- old) grown in the Experimental Farm at El-Qanater El-Khairia Horticultural Research Station (EHRS), Agricultural Research Centre (ARC), Qaloubiya governorate during 2021/2022 and 2022/2023. Disease incidence and disease severity were higher during 2022/2023 than 2021/2022 under natural field infection. Alternaria alternata (Fr.)Keissler and Monilinia fructicola G.Wint. fungi were isolated from diseased inflorescences, diseased spotted leaves and fruits furthermore, Fusarium solani Mart. fungus was isolated from spotted leaves. M. fructicola showed the highest frequency among fungi isolated from diseased inflorescences and leaves but A. alternata was the highest isolated fungus from rotted fruits. Pathogenicity test showed that A. alternata and M. *fructicola* were able to induce inflorescence blight, leaf spot and fruit rot diseases on loquat. On the other hand, little spots were appeared on leaves inoculated by F. solani. In vitro, the efficacy of four agrochemical products (Score 25%, Scar Nat 70%, Pyrmadol 40% and Decent 32.5%) was measured on the linear growth of the isolated fungi. Generally, growth of the isolated fungi was totally inhibited with Score (25% EC), Scar Nat (70% WP), Pyrmadol (40% SC) and Decent (32.5% EC) at the rate of 1ml or 1g/L. Score and Decent were more effective than Scar Nat and Pyrmadol. In vivo, under filed conditions, treatment with Score (100 ml/100 L water) significantly decreased inflorescence blight, leaf spot and fruit rot diseases during the two experimental seasons followed by Decent (100 ml/100 L water) and the lowest efficacy was obtained with Pyrmadol (100 ml/100 L water).

Keywords: Loquat, Eriobotrya japonica. Alternaria alternata, Monilinia fructicola.

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## **INTRODUCTION**

Loquat (Eriobotrya japonica Lindl) is a tropical and subtropical tree within the genus Erobotrya belonging to family Rosaceae, grown commercially for its eatable's fruits. Loquat is the English name but Beshmelah is the Arabic name. Loquat fruit is yellow to orange, contains 3-4 seeds, 5-6 cm long and sweet test as well as pearshaped. Thirty countries *i.e.*, Italy, Turkey, Syria, Egypt and others have planted loquat trees while the trees are native and commercial in China as well as Japan (Alwash, 2017). In Egypt, loquat was introduced at 1805-1849 but the fruit is not widely known. Cultivars grown in Egypt include El-Sukkary, Emanuel and Premiere (Elsabagh, 2011). Loquat trees are liable to infection by several pests, fungal pathogens and other pathogens have been reported to cause diseases on loquat. Moreover, brown rot disease caused by Monilinia fructicola is the most important disease on Loquat and cause many damages on loquat fruit and other parts on trees therefore, worrying disease in the future, the causal pathogen fond and which has been extensively reported on peach, plum, apricot, cherry, apple and pear (Michailides et al., 2007 and Luo, 2017). The first report from China, Monilia spp. were isolated and identified also using molecular analysis from diseased loquat fruits showing fruit rot symptoms. In addition, M. yunnanensis, M. fructicola and M. mumecola cause brown rot disease on loquat fruits. From the three fungi, M. fructicola appeared to be the most virulent one in the pathogenicity on loquat and peach fruits. (Yin et al., 2021). In Japan, Fusarium sp. caused fruit rot disease of loquat (Takushi and Kamekawa 2011). Furthermore, the first report in Pakistan reported that F. solani was isolated from loquat fruit showing rot disease (Abbas et al. 2017). Also, the fungal species in the genus Alternaria are common necrotrophs with different lifestyles, ranging from saprotrophes to endophytes to pathogens. Alternaria alternata causes several diseases on Loquat trees such leaf spot, fruit rot and other diseases in Greece (Tziros, 2013). Furthermore, A. alternata has been reported as the causal pathogen of leaf spot and fruit rot diseases on loquat in Palestine (Batta, 2005). This study focused on loquat inflorescence blight, leaf spot and fruit rot diseases, isolation and identification of the pathogenic fungi. In Egypt, to date many information are missed for about the control of loquat diseases. The present study measured the reduction of inflorescence blight, fruit rot and leaf spot diseases of loquat on Emanuel cultivar (15 years- old), during 2021/2022 and 2022/2023 by using, four agrochemical products, Score 25% EC. Scar Nat 70% WP, Pyrmadol 40% SC and Decent 32.5% EC.

## MATERIALS AND METHODS

# **Disease incidence and disease severity** (%) under natural infection:

This experiment was achieved on loquat trees cv. Emanuel (15-years- old) grown in an experimental farm at El-Qanater El-Khairia, Horticultural Research Station (EHRS), Agricultural Research Centre (ARC), Qaloubiya governorate during 2021/2022 and 2022/2023. Loquat tress was designed as three replicates and each replicate consisted of ten trees. Disease incidence (%) as well as disease severity (%) of (inflorescence blight, fruit rots and leaf spot) were estimated. Disease incidence(%) was calculated using the following formula:

Disease incidence (%) =  $\frac{\text{Number of infected inflorescences, fruits and leaves}}{\text{Total number of inspected inflorescences, fruits and leaves}} \times 100$ The disease severity (%) was determined using the following formula:

$$DS(\%) = \frac{\Sigma n \times v}{5N} \times 100$$

Where:

- n= Number of each of infected inflorescences, fruits and leaves of each grade.
- N= Total number of the inspected inflorescences, fruits and leaves (each alone).
- 5 = Maximum disease severity grade.
- v = Numerical value of each grade as follows: -
  - 0 = healthy inflorescences, fruits and leaves.
  - 1 = above 1 25% of each, inflorescence blight, fruit rots or leaf spot.
  - 2 = above 25- 50% of each, inflorescence blight, fruit rots or leaf spot.
  - 3 = above 50- 75% of each, inflorescence blight, fruit rots or leaf spot.
  - 4 = more than 75% of each, inflorescence blight, fruit rots or leaf spot.
  - 5 = complete inflorescence blight, fruit rots or leaf spot.

## Fungi associated with infected samples:

Isolations were made routinely from symptomatic material of diseased loquat (inflorescence blight, fruit rot and leaf spot). Small pieces of tissues were taken from diseased parts and surface-sterilized in 70% ethanol for 10 sec, 1% NaOCl for 0.5 min, washed two times in sterilized water and dried between two pieces of sterilized filter paper, then planted onto sterilized water agar (WA) medium in Petri plates. Hyphae growing out from the tissue pieces were sub-cultured into sterilized plates containing a new fresh PDA, incubated at 22°C for 3 days. To obtain pure cultures, hyphal-tips were taken to new Potato dextrose agar PDA medium.

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All isolates were preliminary identified according to their cultural characteristics and microscopic examination, at Plant Pathology Research Institute, Agric. Res. Center, using the descriptions of, Booth (1971), Barnett & Hunter (1986) and Simmons (1995). Fungal isolates were grown onto PDA in test tubes maintained at 4°C for further studies. Frequency of the fungal isolates was calculated according to Hassan *et al.*, (2021).

## Pathogenicity test:

Pathogenicity test was carried out during 2021/2022 and 2022/2023 on healthy inflorescences, fruits and leaves of cv. Emanuel, Loquat branch (40 - 50 cm) containing one inflorescence and/or some new leaves were thoroughly washed by water, surface sterilized with 1% sodium hypochlorite solution for one min, followed with washing by sterilized water. Branches with prepared inflorescence and/or leaves were individually inserted into one-liter sterilized glass bottles, each containing nutrient solution (2% K<sub>2</sub>SO<sub>4</sub>, 1% Urea, 0.5% ZnSO<sub>4</sub> and 0.3% Borax) according to Khamis et al. (2007). Concerning fruits, mature loquat fruits were washed by water, surface sterilized with 1% sodium hypochlorite solution for one min, followed by washing with sterilized water. Each inflorescence, leaves and fruits were wounded by small scratch and inoculated individually by spore suspension of Alternaria alternata, Monilinia fructicola and Fusarium solani. each alone

Table (1): Tested agrochemical products.

(10<sup>6</sup> spores/ml) collected from 7:8 days old cultures using atomizer (Lachhab et al., 2015). Nine inflorescences/fungus were used as three replicates and 6 branches each containing 6:9 leaves/ fungus as three replicates. Three replicates for each fungus and 5 fruits were used for each replicate. Loquat fruits were incubated for 10 days at room temperature. Artificially inoculated inflorescences and leaves were kept for 21 days at greenhouse conditions (22:25°C) and assessed for disease symptoms using the disease severity (%) scale as mentioned before. Additionally, the same numbers of unwounded inflorescences, leaves and fruits were inoculated by spraying spore suspension  $(10^6 \text{ spores/ml})$  of Alternaria alternata, Monilinia fructicola and Fusarium solani, each alone. Loquat fruits were incubated for 10 days at room temperature. Artificially unwounded inoculated inflorescences and leaves were kept for 21 days at greenhouse conditions at the same temperature (22:25°C). The causal fungi of inflorescence blight, leaf spot and fruit rot were re-isolated again for identity confirmation.

## **Bioassays of agrochemical products.**

To decrease the percentage of inflorescence blight, fruit rots and leaf spot, four commercial agrochemical products (Table 1) were evaluated *In vitro* on the linear growth of the isolated fungi, and *In vivo* for their effect on loquat trees cv. Emanuel (15-years- old).

= = = = = = = = = = = = = = = = = = = =	ugroenement products.			
Product	Active ingredients	Status	Conc./ L	
Score 25%	Difenoconazole	EC	0.5 and 1ml	
Scar Nat 70%	Thiophanate-methyl	WP	0.5 and 1g	
Pyrmadol 40%	Pyrimethanil	SC	0.5 and 1ml	
Decent 32.5%	Azoxystrobin 20% + Difenoconazole 12.5%	EC	0.5 and 1ml	

## In vitro experiment:

Four agrochemical products, score 25% EC, Scar Nat 70% WP, Pyrmadol 40% SC and Decent 32.5 % EC were evaluated *in vitro* for their efficacy on linear growth of *M. fructicola* and *A. alternata* (the famous causals of inflorescence blight, fruit rots and leaf spot). Treatments were added each

alone to 250 mm flasks containing autoclaved PDA media before solidifying, then poured in 9cm  $\emptyset$  plates. After solidification, the plates were inoculated at the center with disks 5 mm  $\emptyset$  of 7days old cultures of the tested fungus, then incubated at 22:24°C. Untreated plates inoculated with the tested fungi served as control. Six plates were used for each treatment as replicates. When any of the control plates were full growth, the average colony diameter was calculated according to Hassan *et al.*, (2021).

## Field experiment.

This experiment was conducted in a 15vears old cv. Emanuel loquat orchard located in EHRS, ARC, during 2021/2022 and 2022/2023 to evaluate the efficacy of the previously mentioned agrochemical products for controlling inflorescence blight, fruit rot and leaf spot diseases under field conditions using the concentration that gave the best inhibition of the tested pathogens growth in vitro, 1ml or 1g/liter water. Nine trees were used as three replicates for each treatment. Additionally, nine trees sprayed with water only were used as control. Treatments were sprayed three times at 15 days intervals, the first application of the tested agrochemical products was sprayed at fruit set (Dec. 10<sup>th</sup> and 15<sup>th</sup> for 2021 and 2022, respectively). And also, agrochemical products were sprayed twice, 25 and 15 days before harvest. Loquat trees were fertilized, irrigated and sprayed to prevent insect injury for a healthy growth in the orchard. Disease severity levels (%) of inflorescence blight, fruit rot and leaf spot were calculated at fruit mature stage (March 20<sup>th</sup> and 25<sup>th</sup> for 2022 and 2023, respectively). The efficacy of the tested products was calculated using the formula:

$$\% \text{ Efficacy} = \frac{\text{Control}(c) - \text{Trreatment}(t)}{\text{Control}(c)} \times 100$$

Where:

- c= Disease severity of each of inflorescence blight, fruit rot or leaf spot % in the control.
- t= Disease severity of each of inflorescence blight, fruit rot or leaf spot % in treated trees.

#### Statistical analysis:

All the obtained data during both years were subjected to analysis of variance method according to Snedecor and Cochran (1990). Duncan's Multiple Range test (Duncan, 1955) was used to compare differences among means.

# **Disease incidence and disease severity** (%) under natural infection:

Data in Table (2) and Figs. (1, 2 and 3) show that values of disease incidence (DI %) and disease severity (DS %) of loquat inflorescence blight, leaf spot and fruit rot under natural infection were higher during 2022/2023 than 2021/2022. Fruit rot showed the highest averages of disease severity in first and second season. Leaf spot showed the lowest disease incidence and severity in the first and second seasons. Inflorescence blight showed the highest averages of disease averages of disease incidence and severity in the first and second seasons.

Table(2): Disease incidence and disease<br/>severity of loquat inflorescence blight,<br/>leaf spot and fruit rot under natural<br/>infection during 2021/2022 and<br/>2022/2023 seasons.

Diseases	2021/2	2022	2022/2023					
Diseases	DI%	DS%	DI%	DS%				
Inflorescence	36.66a	16b	40a	18b				
blight								
Leaf spot	16.66c	10c	20c	12c				
Fruit rot	30b	20a	33.33b	23a				

\*No significant differences between the treatments with the same letter/s in the same column at  $P \le 0.05$ DI%=Disease incidence & DS%= disease severity

#### Fungi associated with infected trees:

Pure cultures of the isolated fungi were identified as Alternaria alternata (Fr.) Keissler, Monilinia fructicola G.Wint. and Fusarium solani Mart. based on the cultural characteristics and microscopic observations. Data in Table (3) show that the three isolated fungi were associated with loquat inflorescence blight, leaf spot and fruit rot symptoms. A. alternata, M. fructicola were isolated from infected inflorescences, fruits leaves and furthermore, F. solani was isolated only from infected leaves. M. fructicola was the highest isolated fungus from plant organs infected by inflorescence blight and leaf spot but A. alternata showed the highest frequency among the isolated fungi from rotted fruits during the two years of the investigation.

## RESULTS



Fig. (1): Loquat leaves showing natural leaf spot symptoms (A), (B) and (C).



**Fig. (2):** Healthy loquat inflorescence (A), inflorescence showing typical inflorescence blight symptoms (B) and (C).



Fig. (3): Loquat fruit rot symptoms under field conditions. Naturally infected fruit (A), natural typical fruit rot symptoms (B), rotted fruits (C), completely rotted and mummified fruits (D).

#### **Pathogenicity test:**

Data concerning the pathogenicity test show that *A. alternata* and *M. fructicola* were able to induce inflorescence blight, leaf spot and fruit rot diseases on loquat with different degrees (Figs. 4, 5, 6 and 7). On the other hand, little spots were appeared on leaves inoculated by *F. solani*. Additionally, data in Table (4) show that the highest disease severity of each of inflorescence blight, leaf spot and fruit rot on loquat cv. Emanuel during 2021/2022 and 2022/2023 was obtained from the infection by *M. fructicola* followed by *A. alternata*. Data also indicated that values of disease severity during 2021/2022 were highest than those estimated during 2022/2023 under artificial inoculation in pathogenicity tests. Moreover, little amounts of the diseases were appeared on the unwounded inoculated leaves,

inflorescences and fruits by using spore suspension of each of *Alternaria alternata* and *Monilinia fructicola* during or after the incubation period (Fig.8). No symptoms of the diseases were appeared on the unwounded leaves, inflorescences and fruits when inoculated by spore suspension of *Fusarium solani* after the incubation period.

**Table (3):** % Frequency of fungi isolated from loquat trees infected by inflorescence blight,leaf spot and fruit rot diseases during 2021/2022 and 2022/2023.

	Frequency % with							
Isolated fungi	Inflorescence blight		Leaf spot		Fruit rot			
	2021/2022	2022/2023	2021/2022	2022/2023	2021/2022	2022/2023		
Alternaria alternata	48.15b	40.74b	44.44b	37.04b	55.56a	51.85a		
Monilinia fructicola	51.85a	59.26a	48.15a	55.55a	44.44b	48.15b		
Fusarium solani	0.0	0.0	7.41c	7.41c	0.0	0.0		
Total	100	100	100	100	100	100		

\*No significant differences between the treatments with the same letter/s in the same column at  $P \le 0.05$ 

**Table (4):** Disease severity (%) of inflorescence blight, leaf spot and fruit rot caused by thetested fungi, under artificial inoculation during 2021/2022 and 2022/2023 onloquat cv. Emanuel.

			Disease seve	erity (%) on		
Isolated fungi	Inflorescence blight		Leaf spot		Fruit rot	
	2021/2022	2022/2023	2021/2022	2022/2023	2021/2022	2022/2023
Alternaria alternata	34.00b	26.00b	16.00b	14.00b	38.00b	33.00b
Monilinia fructicola	49.00a	44.00a	22.00a	20.00a	48.00a	40.00a
Fusarium solani	0.0	0.0	8.00c	6.00c	0.0	0.0
Control	00c	00 c	00d	00d	00c	00c

\*No significant differences between the treatments with the same letter/s in the same column at P≤0.05



Fig. (4): Healthy loquat inflorescence (A); Symptoms caused by *Alternaria alternata* infection (B) and by *Monilinia fructicola* infection (C) on artificially inoculated loquat inflorescence.



**Fig. (5):** Healthy loquat leaves (A); Symptoms caused by *Alternaria alternata* infection (B) and by *Monilinia fructicola* infection (C) on artificially inoculated loquat leaves.



**Fig. (6):** Symptoms caused by *Alternaria alternata* on artificially inoculated loquat fruits (A, B and C) and control (D).



Fig. (7): Symptoms caused by *Monilinia fructicola* on artificially inoculated loquat fruits (A) and control (B).



**Fig. (8):** Little amounts of the disease on the unwounded leaves, inflorescences and fruits after inoculation by spore suspension using each of *Alternaria alternata* (A, B and C) and *Monilinia fructicola* (D, E and F) after the incubation period.

# Evaluation of the tested agrochemical products:

The efficacy of four agrochemical products namely, score 25% EC, Scar Nat 70% WP, Pyrmadol 40% SC and Decent 32.5 % EC was tested *in vitro* to evaluate their effect on the growth of the tested fungi as well as their active ingredient was used *in vivo* to control loquat inflorescence blight, leaf spot and fruit rot diseases.

#### In vitro test:

Data in Table (5) reveal that all tested treatments reduced the linear growth of M.

*fructicola* and *A. alternata* compared with control. Increasing the concentration from 0.5 to 1 ml or 1g of the treatments increased their effects in inhibiting the growth of the tested pathogens. Generally, a complete reduction in colony growth of the tested fungi was observed at the higher concentration (1ml or 1g /L of any tested agrochemical treatment). The highest reduction was recorded by a score of 25% EC and Decent 23.5% EC compared to Scar Nat 70% WP and Pyrmadol 40% SC.

**Table (5):** Efficacy of four commercial agrochemical products on the linear growth of the isolated fungi *in vitro*.

Treatment	Con /I	Monilinia fru		ructicola Alternaria alternata		Mean	
	Con./L	GR (mm)	RD%	GR (mm)	RD%	GR (mm)	RD%
	0.5 ml	8.0d	91.11	8.0d	91.11	8.0d	91.11
Score 25% EC	1 ml	00.0e	100	00.0e	100	00.0e	100
Scar Nat 70% WP	0.5 g	10.0c	88.88	12.0c	86.66	11.0c	87.77
	1 g	00.0e	100	00.0e	100	00.0e	100
Drymmadal 400/ SC	0.5 ml	13.0b	85.55	15.0b	83.33	14.0b	84.44
Pyrmadol 40% SC	1 ml	00.0e	100	00.0e	100	00.0e	100
Decent 32.5% EC	0.5 ml	8.0d	91.11	8.0d	91.11	8.0d	91.11
	1 ml	00.0e	100	00.0e	100	00.0e	100
Control	-	90a	-	90a	-	90a	-

\* No significant differences between the treatments with the same letter/s in the same column at P $\leq$ 0.05. GR= growth & RD= reduction

#### In vivo experiment:

Efficacy of the tested agrochemical products was evaluated for the control of loquat inflorescence blight, leaf spot and fruit rot diseases under field conditions. Data in Tables (6 and 7) show that 1ml/ L water of Score 25% EC was the best treatment that recorded efficacy 69.99, 68.00 and 75.00 and 80.00, 69.99 and

72.09 % against inflorescence blight, leaf spot and fruit rot during 2021/2022 and 2022/2023 respectively, followed by 1ml/ L Decent 32.5% EC that recorded efficacy 65.00, 63.98 and 70.00 and 76.70, 65.00 and 67.44 % against inflorescence blight, leaf spot and fruit rot during 2021/2022 and 2022/2023 seasons, respectively.

Table (6): Efficacy of four commercial agrochemical products on disease severity (%) of
loquat inflorescence blight, leaf spot and fruit rot diseases, during 2021/2022 season.

		2021/2022						
Treatment	Con./L	Inflorescence blight		Leaf spot		Fruit rot		
		DS%	Eff.%	DS%	Eff.%	DS%	Eff.%	
Score 25% EC	1ml	8.00d	69.99	5.33e	68.00	6.00e	75.00	
Scar Nat 70% WP	1g	13.33b	50.00	6.66c	60.02	9.60b	60.00	
Pyrmadol 40%SC	1ml	13.33b	50.00	7.00b	57.98	8.00c	66.66	
Decent 32.5% EC	1ml	9.33c	65.00	6.00d	63.98	7.20d	70.00	
Water		26.66a		16.66a		24.00a		

\*No significant differences between the treatments with the same letter/s in the same column at P $\leq$ 0.05 DS =Disease severity & Eff. = Efficacy of tested product

The lowest efficacy was recorded when loquat trees were sprayed with 1ml/ L Pyrmadol 40% SC. In general, all the tested commercial agrochemicals reduced

inflorescence blight, leaf spot and fruit rot diseases compared with control treatment in both seasons.

Table (7): Efficacy of four con	mmercial agrochemicals products on disease severity (%) of
loquat inflorescence	blight, leaf spot and fruit rot diseases, during 2022/2023
season.	
	2022/2023

		2022/2023					
Treatment	0 7	Inflorescence blight		Leaf spot		Fruit rot	
	Con./L	DS%	Eff.%	DS%	Eff.%	DS%	Eff.%
Score 25% EC	1ml	4.00e	80.00	4.00d	69.99	4.80e	72.09
Scar Nat 70% WP	1g	5.33c	73.35	5.33b	60.01	6.00c	65.11
Pyrmadol 40%SC	1ml	6.60b	67.00	5.33b	60.01	7.20b	58.13
Decent 32.5% EC	1ml	4.66d	76.70	4.66c	65.00	5.60d	67.44
Water		20.00a		13.33a		17.20a	

\* No significant differences between the treatments with the same letter/s in the same column at P < 0.05DS =disease severity & Eff. = Efficacy of tested product

### DISCUSSION

The fruit of the subtropical loquat tree (Eriobotria japonica Lindl.) blossoms in the fall, grows throughout the winter, and ripens in the spring before any other fruit, which closes a market gap. Because there aren't many competing fruits on the market in the spring, loquat fruits can be sold for more money (Sultan et al., 2015). In addition to being consumed raw, loquat is currently used in the production of jam and jellies (Koba et al., 2007; Delfanian, 2015 and Alwash, 2017). In China and Japan, loquat is widely recognized as a medicinal plant (Fouedjou et al., 2016) and has been a part of traditional medicine for a millennium (Alwash, 2017). While, loquat fruits are perishable with a postharvest shelf life about 10 days at room temperature because of decay and browning rot which causes economic losses during handling and storage or in marketing (Cao et al., 2014). In the present study, results indicated that disease incidence and disease severity of loquat inflorescence blight, leaf spot and fruit rot diseases under natural infection were higher during 2022/2023 than 2021/2022. Fruit rot and inflorescence blight showed the highest average of disease severity in first and second seasons. Leaf spot showed the lowest disease incidence and severity

in first and second seasons. Also, in this study Alternaria alternata and Monilinia fructicola were isolated from plant material infected by inflorescence blight, leaf spot and fruit rot. Furthermore, Fusarium solani was isolated only from diseased leaves. M. fructicola was the highest fungus isolated from inflorescence blight and leaf spot but A. alternata was the highest isolated fungus from rotted fruits during the two vears of the present investigation. Alternaria alternata and M. fructicola were able to induce inflorescence blight, leaf spot and fruit rot diseases on loquat with different degrees. On the other hand, little spots were appeared on leaves inoculated by F. solani. The highest values of disease severity due to inflorescence blight, leaf spot and fruit rot on loquat cv. Emanuel were obtained from those infected by *M. fructicola* followed by those infected by A. alternata. The obtained results are consistent with Cao et al., (2014) Numerous reports have indicated that Monilinia fructicola is the cause of brown rot disease on a wide variety of fruits across the globe. On peaches, apricots, apples, pears, and loquats, reports have been made. Serious losses have been reported in Europe, Asia, and South America due to brown rot on apples, pears, and stone fruit caused by Monilinia sp.

(Ogawa and English, 1991). Snowdon (1990) reported that Alternaria alternata caused serious damage on pears in Korea and Japan. Li et al., (2007) reported that A. alternata can cause fruit rot on pear in China. A. alternata can infect fruits before or after harvest through weak or injured tissue associated with chemical or mechanical injury (Jones and Aldwinckle, 1990). Many reports are published about Alternaria sp. causing leaf spot and fruit rot on loquat in Florida, Japan, Mexico and Taiwan (Ko et al., 2010 and Farr and Rossman, 2013). In Iran, Brown rot and leaf spots of loquat trees are caused by Monilinia fructigena and Spilocea pyracanthae, respectively (Ershad, 2009). A. alternata was isolated from loquat leaves and fruits, and its pathogenicity was confirmed (Mirhosseini et al., 2015). Our results showed that all tested treatments reduced the linear growth of *M. fructicola* and A. alternata on PDA media compared with control. The highest reduction was recorded by Score 25% EC and Decent 23.5 % EC compared to Scar Nat 70% WP and Pyrmadol 40% SC. In vivo experiment 1ml/ L water of Score 25% EC was the best treatment followed by 1ml/ L Decent 32.5 EC. While the lowest efficacy was recorded when loquat trees were sprayed with 1ml/ L Pyrmadol 40% SC. In general, treatments reduced infection all bv inflorescence blight, leaf spot and fruit rot compared with control treatment in both investigating seasons. These results are in agreement with those recorded by Batta (2005)who found that Score (Difenoconazole) and Switch (cyprodinil+flodioxonil) significantly reduced the lesion diameter of A. alternata on treated loquat fruits and leaf discs compared to the control fruits and leaf discs. Also, application these compounds completely inhibited the sporulation of A. alternata compared to the untreated control. Chemical fungicides were significantly more effective than the biofungicides, and all the tested compounds particularly Score, 25% SC (Difenoconazole), Amistar 25% SC (Azoxystrobin) and Ridomil 68% WP

(Metalaxyl + Mancozeb) significantly reduced incidence and severity of early blight and increased fruits in tomato yields in comparison with control (El-Kholy et al., 2021). Score25% SC (Difenoconazole) and Eminent (Tetraconaeole) were the most effective commercial products in controlling leaf spot of sugar beet (El-Sayed et al., 2017). Score 25% EC (Difenoconazole 25%) and Bellis 38% WG (25.2% boscalid+12.8% pyraclostrobin) had more effect on apple scab infections and yield productivity (kg/tree), followed by Occidor 50%SC (Carbendazim 50%) and Topsin M-70WP (Thiophanate- methyl 70%)(Radwan and Hassan. 2019). Benzimidazole group is a systemic fungicides such as (Thiophanate-methyl) against a lot of fungal diseases (Morsy and EL-Hefny 2017). Azoxystrobin at 2 and 4 ml/l significantly reduced the development of both panicle and leaf anthracnose of mango trees in field experiment (Sundravadana et al., 2007). Hassan et al., (2024) found that, 0.5ml/L Difenoconazole 25 % (Score 25% EC), 0.5g/L Boscalid 25.2 % + Pyraclostrobin 12.8% (Bellis 38%WG) and 1ml/L Thiabendazole (Tekto 50 % SC) completely inhibited the linear growth of all the isolated fungi. Under filed conditions Score 25% EC gave the highest efficacy for controlling fruit drop of mango during the two seasons of investigation on both tested cultivars cv. Montakhab El-Qanater and cv. Ewais, followed by Bellis 38% WG and Tekto While 40 ml/100L 50% SC. water Difenoconazole 15% + Propiconazole 15% (Craft 30% EC) and 25ml/100 L water Azoxystrobin 20% +Tebuconazole 30 % (Dovex 50% SC) gave the lowest efficacy.

## CONCLUSIONS

This study suggests that *Monilinia fructicola* and *Alternaria alternata* are the main pathogens causing loquat inflorescence blight, leaf spot and fruit rot diseases during 2021/2022 and 2022/2023 in Egypt. Artificial inoculation using these fungi induced typical disease symptoms. Also, loquat trees pre-harvest sprayed three times with 15 days intervals using the

tested commercial products, the first application was sprayed at fruit set and also treatments were sprayed twice, 25 and 15 days before harvest with the best concentration of Score 25% EC (Difenoconazole) 1ml/litre, Scar Nat 70% WP (Thiophanate-methyl) 1g / litre, Pyrmadol 40% SC (Pyrimethanil)1ml/litre and Decent 32.5EC (Azoxystrobin 20% + Difenoconazole 12.5%)1ml/litre may help in the control of loquat of inflorescence blight, leaf spot and fruit rot diseases.

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## **REFERENCES:**

- Abbas, M. F.; Naz, R. F.; Zhang, C. A. X.; Rosli, B. H.; Gleason, M. L. and Mehmood, N. (2017). First report of *Fusarium solani* causing fruit rot of loquat (*Eriobotrya japonica* Lindl) in Pakistan. Plant Disease, 1-2 p.
- Alwash, B. M. J. (2017). Cytotoxic and antioxidant activity of fruit juice of *Eriobotrya Japonica* (Thunb.) Lind. Plant cultivated in Iraq. The Iraqi Journal of Agricultural Sciences; 48(3): 892-898.
- Batta, Y. (2005). Control of *Alternaria alternata* on loquat (*Eriobotrya japonica* Lindl.) using detached fruits and leaf-disk assay. An-Najah Univ J Res (N Sc) 19:69-82.
- Barnett, H. L. and Hunter, B. B. (1986).
  Illustrated Genera of Imperfect Fungi.
  4<sup>th</sup> Ed., Macmillan Publishing Co., New York, 218 pp.
- Booth, C. (1971). The Genus Fusarium. Kew, Surrey. Commonwealth Mycol. Inst., 2nd Ed., 237pp.
- Cao, S.; Cai, Y.; Yang, Z.; Joyce, D. C. and Zheng, Y. (2014). Effect of MeJA treatment on polyamine, energy status and anthracnose rot of loquat

fruit. Food Chem 145, 86-89. https://doi.org/10.1016/j.foodchem.08. 019.

- Delfanian, M.; Kenari, R. E. and Sahari, M. A. (2015). Antioxidant activity of loquat (*Eriobotrya japonica* Lindl.) fruit peel and pulp extracts in stabilization of soybean oil during storage conditions. International Journal of Food Properties; 18: 2813-2824.
- **Duncan, D. B. (1955).** Multiple range and multiple F. tests. Biometrics, 11: 1-42.
- El-Kholy, R. M.; El-Samadesy, A. M.; Helalia, A. A. and El-Ballat, E. M. (2021). Efficacy of certain chemical fungicides and biofungicides on early blight disease in tomato under field conditions. Al-Azhar Journal of Agricultural Research V. (46) No. (2): 145-153.
- El-Sayed, A. B. B.; El-Shehawy A. E.;
  El-Shabrawy, E. M. and Alkolaly A.
  M. (2017). Safely application of some fungicides to control cercospora leaf spot disease of sugar beet. Egypt. J. Phytopathol., Vol. 45, No. 2, 219-230 pp.
- Ershad, D. (2009). Fungi of Iran. Iranian Research Institute of Plant Protection, Tehran. pp.531
- Elsabagh, A. S. (2011). Production of Loquat in Egypt. Acta. Hort, 887: 123-126.
- Farr, D. F. and Rossman, A. Y. (2013). Fungal databases, systematic mycology and microbiology laboratory, ARS, USDA. Retrieved March 11, 2013, from http://nt.arsgrin.gov/fungaldatabases/
- Fouedjou, R. T.; Nguelefack-Mbuyo, E.
  P.; Ponou, B. K.; Nguelefack, T.B.;
  Barboni, L. and Tapondjou, L. A.
  (2016). Antioxidant activities and chemical constituents of extracts from *Cordyline fruticosa* (L.) A. Chev. (Agavaceae) and *Eriobotrya japonica* Lindl, (Rosaceae). Pharmacologia; 7 (2-3): 103-113.
- Hassan, M. S. S.; Monir, G. A. and Radwan, M. A. (2021). Efficacy of certain essential oils, copper oxide,

copper oxide nanoparticle, Imazalil and *Bacillus subtilis* to control fruit rot of avocado. Egypt. J. Phytopathol., 49(1): 166-181.

- Hassan, M. S. S.; Shehata, A. S. F. and Banora, M. Y. (2024). Impact of some agrochemical products on early fruit drop of certain Egyptian mango cultivars induced by fungal infection. Egyptian Journal of Phytopathology, Vol. 52, No.1, pp 67-82.
- Jones, A. L. and Aldwinckle, H. S. (1990). Compendium of Apple and Pear Diseases. APS Press, Saint Paul, MN. 224 p.
- Koba, K.; Matsuoka, A.; Osada, K. and Huang, Y. S. (2007). Effect of Loquat (*Eriobotrya japonica*) extracts on LDL oxidation. Food Chemistry; 104(1): 308-316.
- Ko, Y.; Liu, C. W.; Chen, S. S.; Chen, C.
  Y.; Yao, K. S.; Maruthasalam, S.
  and Lin, C.H. (2010). First report of fruit rot of loquat caused by an *Alternaria* sp. in Taiwan. Plant Dis 97(4):481.
- Khamis, M. A.; Bakry, K. A. and Abd El-Moty, S. A. (2007). Improving growth and productivity of guava trees. Minia J. Agric. Res. Dev., 27, 51-70.
- Lachhab, N.; Sanzani, S.M.; Fallanaj,
  F.; Youssef, K.; Nigro, F.; Boselli,
  M. and Ippolito, A. (2015). Protein hydrolysates as resistance inducers for controlling green mould of citrus fruit. Acta Hortic., 1065: 1593-1598.
- Li, Y.; Bi, Y. and An, L. (2007). An occurrence and latent infection of Alternaria rot of Pingguoli pear (*Pyrus bretschneideri* Rehd. cv. Pingguoli) fruits in Gansu, China. J. Phytopathol. 155(1):56-60.
- Luo, C. X. (2017). Advances and prospects on researches of brown rot disease on fruits. Acta Phytophylacica Sinica, (In Chinese) 47, 145-153.
- Michailides, T. J.; Luo, Y.; Ma, Z. H. and Morgan, D. P. (2007). Brown rot of dried plum in California, new insights on an old disease. APSnet Features,

doi: 10.1094/APSnetFeature-0307.

- Mirhosseini, H. A.; Babaeizad, V. and Basavand, E. (2015). Identification and detection of agent of loquat leaf spot and fruit rot in north of Iran. Journal on New Biological Reports. JNBR 4(2) 135 - 138.
- Morsy, A. R. and EL-Hefny, D. E. (2017). Residues assessment of Captan, Spirodiclofen and Thiophanate methyl in Apple fruits under the field conditions. Middle East J. Agric. Res., 6(1): 135-142,
- Ogawa, J. M. and English, H. (1991). Diseases of Temperate Zone Tree Fruit and Nut Crops,. UCANR Publications, Oakland, CA. 464 p.
- Radwan, M. A. and Hassan, M. S. S. (2019). Determination of the optimizing fungicidal applications for controlling apple scab disease in Egypt. Menoufia J. Plant Prot., Vol. 4 June: 69 82.
- Simmons, EG. (1995). Alternaria themes and variations (112-144). Mycotaxon 55:55-163
- Snowdon, A. L. (1990). Pome fruits. In A Colour Atlas of Postharvest Diseases and Disorders of Fruits and Vegetables, Volume 1. Wolfe Scientific, London, UK. 170-216.
- Sundravadana, S.; Alice, D.; Kuttalam, S. and Samiyappan, R. (2007). Efficacy of azoxystrobin on *Colletotrichum gloeosporiodes* growth and on controlling mango anthracnose. Journal of Agricultural and Biological Science. 2(3):10-15.
- Snedecor, G.W. and Cochran, G.W. (1990). Statistical Methods. 8th Ed. The Iowa State Univ., Press Ames, Iowa. U.S.A. 503-507 pp.
- Sultan, M. Z.; Khalefa, S. M.; Elhamamsy, S. M. and Mostafa, Y. S. (2015). Effect of postharvest antioxidant treatments on loquat fruit deterioration during storage at room temperature. Acta Hortic. 1092, 173-179.

https://doi.org/10.17660/ActaHortic. 2015.1092.26.

- **Takushi, T. and Kamekawa, A. (2011).** Okinawa Prefectural Agricultural Research Center Report 5:1.
- **Tziros, G. T. (2013).** *Alternaria alternata* causes leaf spot and fruit rot on loquat (*Eriobotrya japonica*) in Greece. Australasian Plant Dis. Notes, 8:123-124
- Yin, Liang-fen; Zhang, Shu-qin; DU, Juan; WANG, Xin-yu; XU, Wenxing and LUO, Chao-xi (2021). *Monilinia fructicola* on loquat: An old pathogen invading a new host. Journal of Integrative Agriculture, 20(7): 2009-2014.



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